Metal Coordination-Directed Hydroxylation of Steroids with a Novel Artificial P-450 Catalyst

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A novel catalyst has been synthesized in which a manganese-porphyrin unit is linked to two 2.2'-bipyridyl groups and two pentafluorophenyl groups in trans fashion on its four meso positions. Relative to a previous catalyst in which the manganese-porphyrin had four 2,2'-bipyridyl groups, the new catalyst, in the presence of Cu²⁺ ions as coordinating linkers, catalyzes the oxidation of a steroid substrate with much better regioselectivity and higher turnover numbers.

We have described hydroxylation of steroids by mimics of the enzyme cytochrome P-450 class. In our earliest examples, we used metalloporphyrins carrying cyclodextrin groups to bind hydrophobic substrates in water. When the cyclodextrins were linked to *p*-phenylene groups attached to the porphyrin meso positions in catalyst 1, we saw highly selective hydroxylation at the C-6 α position of a steroid derivative 2 doubly bound into the cyclodextrins (Figure 1).¹ However, there were only about 10 turnovers before the catalyst was itself oxidatively destroyed. When instead the phenylene linkers in the catalyst were replaced by tetrafluorophenylene groups in catalyst 3, we still saw hydroxylation selectively at C-6 α , but now with 187 turnovers before destruction of the catalyst (Figure 1).² That same perfluorinated catalyst was also able to hydroxylate the steroid in the C-9 alpha

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position when a substrate was hydrophobically bound to three cyclodextrins of the catalyst.³

There could be advantages in changing the mode of substrates/catalyst binding. Hydrophobic binding is somewhat ill-defined geometrically, and it is seen only in water solvent. We had used double metal-ion coordination to bind a substrate to a metalloporphyrin and a metallosalen, and had observed good selective epoxidation of the substrate.⁴ Thus, we used manganese porphyrin 4 to bind and oxidize substrates, e.g., 5, in the presence of coordinating metals such as Cu²⁺ ions (Figure 2).⁵ However, the oxidation was not selective, with observed formation of six almost evenly distributed products, including 6. Moreover, the turnover number was only 5, low compared to the 187 turnovers we observed with catalyst 3.

The poor catalytic performance in this biomimetic study might result from two factors-the intrinsic reactivity and



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Figure 1. Selective hydroxylation by a catalyst with attached cyclodextrins.

the oxidative stability of catalyst 4 are significantly lower compared with catalyst 3, and the nitrogen atoms from



Figure 2. Metal coordination-directed oxidation.

both the manganese porphyrin and the substrate are easily oxidized to *N*-oxides under the hydroxylation conditions.⁶ Once *N*-oxides are formed, the nitrogens lose the ability to coordinate to the metal center. Indeed, we isolated an initial undesired substrate oxidation product, the bis *N*-oxide of **5** from the hydroxylation experiments before basic hydrolysis;⁷ this was not further hydroxylated under the same conditions.

To avoid the undesired substrate oxidation, we attached different metal ligands to the steroid, two α -phosphonoacetyl groups in substrate **7** (Figure 3).^{8,9} When **7** was subjected to



Figure 3. Hydroxylation of substrate 7 with catalyst 4.

hydroxylation with iodosobenzene and the manganese porphyrin **4** in a mixture of H₂O and *t*-BuOH (1:1), products resulting from hydroxylation on the 5 α , 6 α , 7 α , 7 β , 12 α , and 14 α positions were identified by ¹H NMR.^{10,11} Ten catalytic turnovers were observed.

To further address the poor regioselectivity as well as the low turnover in the metal coordination-directed oxidation, we designed a new catalyst **8** with the introduction of two C_6F_5 groups on its meso positions.

(7) ¹H NMR (300 MHz, CDCl₃): δ = 8.80 (d, 2H, *J* = 6.0 Hz), 8.37 (d, 2H, *J* = 6.2 Hz), 7.90 (d, 2H, *J* = 7.8 Hz), 7.40 (t, 2H, *J* = 6.5 Hz), 4.97 (m, 1H, C3α-H), 4.86 (t, 1H, *J* = 7.7 Hz, C17α-H), 0.91 (s, 1H, C19-Me), 0.84(s, 1H, C18-Me). CI-MS: 535.3 [M + H]⁺.

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(9) Compound 7 was synthesized from the commercially available 3β , 17β -androstanediol and 2-dibenzylphosphonate acetic acid in a three-step procedure. ¹H NMR (400 MHz, D₂O): $\delta = 4.47$ (m, 1H, C3 α -H), 4.40 (t, J = 8.0 Hz, 1H, C17 α -H), 2.55–2.41 (dd, 4H), 0.670 (s, 3H, C19-Me), 0.639 (s, 3H, C18-Me).

(10) The chemical shifts of C19 and C18 angular methyl protons in the identified steroid products (CDCl₃, 400 MHz). 3β ,17 β -Androstanediol (starting material): $\delta = 0.819$ (C19-Me), 0.732 (C18-Me). 3β ,5 α ,17 β -Androstanetriol: $\delta = 1.003$ (C19-Me), 0.738 (C18-Me), calcd 1.00, 0.73. 3β ,6 α ,17 β -Androstanetriol: $\delta = 0.833$ (C19-Me), 0.738 (C18-Me). Identical to authentic sample; 3β ,7 α ,17 β -androstanetriol: $\delta = 0.823$ (C19-Me), 0.738 (C18-Me). Identical to (C18-Me); calcd 0.81, 0.74. 3β ,7 β ,17 β -Androstanetriol: $\delta = 0.823$ (C19-Me), 0.763 (C18-Me), calcd 0.85, 0.76; 3β ,14 α ,17 β -Androstanetriol: $\delta = 0.844$ (C19-Me), 0.864 (C18-Me), calcd 0.82, 0.85.

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Scheme 1



Compared to the previous manganese porphyrin catalyst **4**, the new artificial P-450 catalyst **8** brought the following advantages: (1) The incorporation of two strongly electron-withdrawing pentafluorophenyl groups (C_6F_5) increased the enzyme's reactivity and oxidative stability, and (2) the "*trans*" geometrical relationship between the two bipyridyl ligands eliminates the possibility of undesired *cis*-binding between the artificial P-450 enzyme and the substrate, which may be the cause of some of the side reactions with catalyst **4**.

The synthesis of catalyst 8 was first attempted using either a statistical approach under standard Alder-Longo conditions¹² or an approach developed by Lindsey toward the synthesis of trans-A2B2-porphyrins prepared by reaction of an aldehyde and a dipyrromethane.¹³ However, these two attempts failed. Finally, 8 was successfully synthesized following a Negishi-coupling approach (Scheme 1). The dipyrromethane¹⁴ and the commercially available pentafluorobenzaldehyde were condensed under acid conditions followed by DDQ oxidation to form a porphyrin which was immediately protected as the zinc complex 9.15 Then 9 was regioselectively brominated with NBS16 to give compound 10.¹⁷ which was coupled with a bipvridvl zinc bromide¹⁸ under Negishi conditions to give zinc porphyrin 11.¹⁹ This was demetalated with trifluoroacetic acid to give porphyrin 12,²⁰ which was then metalated with $MnCl_2$ in air to give catalyst 8.21

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- (15) ¹H NMR (400 MHz, acetone- d_6): $\delta = 10.51$ (s, 2H, meso proton),
- 9.61 (d, 2H, J = 4.5 Hz), 9.27 (d, 2H, J = 4.5 Hz). CI-MS: 705.7 [M]⁺.
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- (17) ¹H NMR (400 MHz, acetone- d_6): $\delta = 9.77$ (d, 4H, J = 4.7 Hz), 9.16 (d, 4H, J = 4.7 Hz). CI-MS: 863.0 [M]⁺.
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As hoped, **8** catalyzed the hydroxylation of substrate **7** very regioselectively (90% of the product) at the 6α position with a significantly improved turnover number of 32 (Figure 4). The significant increase in the turnover number is not surprising, considering the nonoxidizable nature of the *substrate*'s coordination groups. The formation of an *N*-oxide of **8**, along with other oxidative degradations, are probably the major factors that limit the turnover number to 32 in the hydroxylation of **7**.



Figure 4. Hydroxylation of substrate 7 with catalyst 8.

Both the selectivity and turnover number of metal coordination directed steroid hydroxylation are not yet quite as

^{(19) &}lt;sup>1</sup>H NMR (300 MHz, CDCl₃): δ = 9.48 (s, 2H), 9.06 (d, 4H, *J* = 4.7 Hz), 8.96 (d, 4H, *J* = 4.7 Hz), 8.87 (m, 4H), 8.70 (m, 4H), 8.00 (m, 2H), 7.51 (m, 2H). FAB-MS: 1013 [M]⁺.

good as the hydrophobic interaction directed steroid hydroxylation by **3**. However, both the regioselectivity and catalytic turnover in the hydroxylation of steroids are probably good enough for synthetic applications. This system may also be more practical since it will not require a water solvent and substrate water solubility for effective catalyst/substrate binding. Acknowledgment. This work has been supported by the NSF and NIH.

Supporting Information Available: Experimental procedures for the syntheses and characterization of the products. This material is available free of charge via the Internet at http://pubs.acs.org.

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^{(20) &}lt;sup>1</sup>H NMR (300 MHz, CDCl₃): δ = 9.50 (s, 2H), 8.99 (d, 4H, *J* = 4.8 Hz), 8.86 (m, 8H), 8.68 (m, 4H), 7.96 (m, 2H), 7.45 (m, 2H), -2.80 (s, 2H, pyrrole NH). MS (FAB): 951 [M]⁺. UV–vis (CH₂Cl₂, nm): 376 (Soret), 437, 470, 504, 544.

⁽²¹⁾ UV-vis (CH₂Cl₂, nm): 331, 435 (Soret), 535, 573.